We drive RNA research with tailored molecular tools

riboPOOL™ Probes Protocol

Ribosomal RNA depletion

Catalog No. dp-P012, dp-P024, dp-P096

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PRODUCT DESCRIPTION

riboPOOLs™ efficiently remove abundant ribosomal RNAs (rRNAs) from total or fragmented RNA prior to downstream analysis by Next-Generation Sequencing or other methods.

riboPOOLs™ are provided either as probes or in a complete kit. **This protocol is for the use of riboPOOL probes alone with reagents from other commercial vendors (as listed below).** For riboPOOL kits, please refer to our riboPOOL kit manual provided.

riboPOOLs™ are available for diverse species and can be tailor-made towards ribosomal RNA or other abundant RNAs from any species. Combination riboPOOLs are also available for rRNA depletion from samples containing multiple species (e.g. for metatranscriptomics or pathogen-infected tissue). riboPOOLs™ consist of complex mixtures of single-stranded 3'- biotinylated DNA probes designed to specifically hybridize with cytoplasmic and mitochondrial rRNAs, enabling their removal with streptavidin-conjugated magnetic beads. The workflow with riboPOOLs™ can be completed in ~70 min (with one-step depletion and RNA clean-up), is enzyme-free, and compatible with high-throughput automation. As it does not rely on polyA-selection, it can be used to detect non-polyadenylated RNA including non-coding RNA, histones and prokaryotic RNA with uniform transcript coverage.

MATERIALS PROVIDED

- riboPOOLs™ (freeze-dried)
- Nuclease-free water

Additional materials and equipment required for rRNA depletion protocol with riboPOOL probes (not provided):

- Streptavidin-coated magnetic beads, such as:
  - **siBeads** from siTOOLs Biotech (streptavidin-coated magnetic beads, 25 mg/ml)
  - Dynabeads MyOne Streptavidin C1 from Thermo Fisher (#65001, 10 mg/ml)
  - Hydrophilic Streptavidin Magnetic Beads from New England Biolabs, NEB (#S1421S, 4 mg/ml)
- Sterile, low-binding 1.5 ml or 2 ml tubes and low-retention tips for minimal surface binding of RNA and beads
- Magnetic tube rack
- Temperature-controlled mixer or thermal cycler
- RNase inhibitor (optional)
- DNase treatment (optional)

Buffers (not provided):
All buffers should be made in DEPC-treated or nuclease-free water.

Buffer Name	Components	Minimum volume required per sample (µl)	Volume for 20 reactions (µl)
Hybridization Buffer (HB)	10 mM Tris-HCl (pH 7.5) 1 mM EDTA 2 M NaCl	5.5	110
Depletion Buffer (DB)	10 mM Tris-HCl (pH 7.5) 1 mM EDTA 1 M NaCl	286	5720
*Bead Resuspension Buffer	0.1 M NaOH 0.05 M NaCl	220	4400
*Bead Wash Buffer	0.1 M NaCl	110	2200

*These buffers are only required when using Dynabeads from Thermo Fisher.

RIBOPOOLS PROBES PROTOCOL

1. **Resuspension of riboPOOL**
   a. Centrifuge tube containing riboPOOL at 11 000 *x g* for 30s before opening.
   b. Resuspend riboPOOL in nuclease-free water provided according to below table (e.g. 15 µl for 12 reaction).

Reaction size	H_2O to add (µl)
12 reaction	15
24 reaction	30
96 reaction	110

2. **Hybridization of riboPOOL to RNA**
   a. To 14 µl of RNA sample (100 ng - 5 µg of total RNA), add:
   If sample volume is > 14 µl, adjust HB volume accordingly to 0.25X total volume. Total volume however should not exceed 40 µl.
      i. 1 µl of resuspended riboPOOL
      ii. 5 µl of **Hybridization Buffer**
      iii. RNase inhibitor (optional)
      Follow manufacturer's instructions for volume required and ensure enzyme is active at 68°C. RNase inhibitor may also be introduced during bead preparation.
   b. Vortex well and spin down droplets.
   c. Incubate at 68°C for 10 min to denature RNA.

d. Allow to **cool slowly** from 68°C to 37°C for optimal hybridization.
To do this, shut off the heating block and let temperature fall naturally to 37°C. If using temperature-controlled ramping, cool at 3°C/min.

3. Preparation of beads

We recommend to perform ribosomal RNA depletion with riboPOOLs in one step. See riboPOOL Application Note for results.

rRNA depletion – bead preparation
If using siBeads by siTOOLs Biotech:

a. Resuspend the siBeads by carefully vortexing tube at medium speed.
b. Transfer 90 µl of bead suspension per sample into a fresh tube.
For batch washing of beads for multiple samples, aliquot bead suspension for up to 6 (i.e. 540 µl) or 12 samples (i.e. 1080 µl) in a single tube.
c. Place tube on magnetic rack and wait for 1 min.
Beads may stick to sides of tube making solution appear brown. Aspirated solution however, should be clear.
d. Aspirate and discard all supernatant.
e. Add 80 µl per sample (i.e. 480 µl for 6 samples, 960 µl for 12 samples) of Depletion Buffer (DB) and agitate the tube well to resuspend beads.
f. Repeat steps c to e.

If using Dynabeads MyOne Streptavidin C1, Thermo Fisher #65001:

a. Resuspend the beads by carefully vortexing tube at medium speed.
b. Transfer 160 µl of bead suspension (10 mg/ml) per sample into a fresh tube.
To prepare multiple samples, aliquot bead suspension for up to 6 samples (i.e. 960 µl) in a single tube.
c. Place tube on magnetic rack and wait for 1 min.
Beads may stick to sides of tube making solution appear brown but aspirated solution should be clear.
d. Aspirate and discard all supernatant.
e. Add 100 µl per sample (i.e. 600 µl for 6 samples) of Bead Resuspension Buffer and agitate the tube well to resuspend beads. Place on magnetic rack (1 min), aspirate and discard supernatant.
f. Repeat step 3e.
g. Add 100 µl per sample of Bead Wash Buffer to resuspend beads. Place on magnetic rack (1 min), aspirate and discard supernatant.
h. Resuspend beads in 80 µl per sample (i.e. 480 µl for 6 samples) of Depletion Buffer.

If using Hydrophilic Streptavidin Magnetic Beads, NEB #S1421S:

a. Resuspend the beads by carefully vortexing tube at medium speed.
b. Transfer 600 µl of bead suspension (4 mg/ml) per sample into a fresh tube.
To prepare multiple samples, aliquot bead suspension for up to 3 samples (i.e. 1800 µl) in a single 2 ml tube.
c. Place tube on magnetic rack (1 min), aspirate and discard supernatant.
d. Add 100 µl per sample (i.e. 600 µl for 6 samples) of Depletion Buffer and agitate the tube well to resuspend beads.
e. Place on magnetic rack (1 min), aspirate and discard supernatant.
f. Resuspend beads in 80 µl per sample (i.e. 480 µl for 6 samples) of Depletion Buffer.

4. **Ribosomal RNA depletion**

a. Briefly centrifuge the tube containing 20 µl hybridized riboPOOL and total RNA (from step 2) to spin down droplets.
b. Combine 80 µl of prepared beads (from step 3f) with 20 µl of the hybridized riboPOOL-RNA solution. Agitate the tube to resuspend well.
c. Incubate at 37°C for 15 min, followed by a 50°C incubation for 5 min.
d. Briefly spin down droplets.
e. Place on magnetic rack for 2 min. Carefully transfer the supernatant into a new tube.
f. Place the new tube with the supernatant on the magnetic rack for 1 min to get rid of trace amounts of beads. (optional)
g. Carefully transfer 90 µl l of the supernatant to a new tube.

Steps f and g are recommended to remove any trace amount of beads.

At this point, RNA can be stored at -20°C overnight or -80°C for up to a month.

5. **Proceed to RNA purification**

RNA samples that have been subject to rRNA depletion **must** be purified before sequencing library preparation to remove salts and buffer concentrates. The following clean-up methods have been successfully applied with riboPOOLs. As some clean-up methods incorporate size selection, please pay careful attention to product specifications and accompanying protocols:

- Ethanol/Isopropanol Precipitation
  Recovers small and large RNAs e.g. tRNAs, miRNAs, mRNA and large non-coding RNA.

- SPRI bead-based RNA clean-up, recommended kits:
  - riboPOOL cleanUP module
  - Agencourt RNAClean XP from Beckman Coulter (#A63987, #A66514)

- Silica column-based RNA clean-up, recommended kits:
  - Clean & Concentrator kit from Zymo Research (#R1013)
    Can be used to recover both small (17-200 nt) and large RNA (> 200 nt) in a single fraction or separate fractions.
  - Nucleospin RNA Clean-up XS from Macharey Nagel (#740903)
    Recovers only large RNA (> 200 nt).

Optional DNase treatment
Our experience and customer feedback supports no riboPOOL contamination after rRNA depletion, which makes a DNase treatment unnecessary. If for another reason DNA treatment is desired, please include a DNase step after RNA clean up, as salt concentration after depletion is too high for efficient DNase treatment. Please include another RNA clean up step after DNase treatment.

ADDITIONAL NOTES

- riboPOOLs work best for high quality RNA samples though can also be used for degraded/ribosome profiling samples.
- RNA input amount may range from 10 ng to 5 µg (ideal range is 100 ng – 1 µg)
- RNA input should be DNA-free.
- Due to abundance of rRNA, expect to lose ~95% of initial input. *Expected yield for 1 µg of input RNA is < 30-50 ng.*
- During protocol, avoid leaving tubes with RNA at room temperature over an extended period of time.

- To agitate beads, flick tube gently till solution becomes homogenous. Alternatively, vortex tube at medium speed.
- Take necessary precautions to avoid RNase contamination.

STORAGE TEMPERATURE

riboPOOLs when lyophilized, are stable at RT for up to a year, but is best stored at -20°C upon receipt. Upon resuspension in nuclease-free water (H_2O), riboPOOLs are stable for at least one year when stored at or below -20°C. Please store in aliquots to avoid freeze-thaw cycles.